LETTERS 2006 Vol. 8, No. 10 2171–2174

ORGANIC

Chemical Transformation of Prostaglandin-A₂: A Novel Series of C-10 Halogenated, C-12 Hydroxylated Prostaglandin-A₂ Analogues

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Received March 16, 2006

ABSTRACT



Synthesis of a novel class of C-10 halogenated and C-12 oxygenated prostaglandin- A_2 derivatives (6a–6c) has been accomplished. (15*S*)-Prostaglandin- A_2 (1), from the gorgonian *Plexaura homomalla*, served as the starting material for the synthesis. The absolute configuration was determined using NMR.

 Δ^{12} -Prostaglandin J₂ (Δ^{12} -PGJ₂), a cross-conjugated enone, is known to inhibit ubiquitin-specific isopeptidase activity causing apoptosis.¹ Mechanistic studies have indicated that inhibition results from Michael addition of an isopeptidase cysteine residue to the endocyclic β -carbon of the cyclopentenone.² Punaglandins (PNGs), C-10 chlorinated and C-12 oxygenated prostanoids, are more potent inhibitors of isopeptidase activity than Δ^{12} -PGJ₂ and the PGA series (Figure 1).³ On the basis of these results, increasing the electrophilicity



Figure 1. Electrophilic prostanoids.

of the endocyclic β -carbon (C-10) should increase reactivity. To test this hypothesis, a new series of halogenated PGA₂ derivatives were synthesized (**4a**-**4c**) by substituting electronwithdrawing groups (Cl, Br, I) at the α -position of PGA₂ (Scheme 1).

Preliminary work with these α -halogenated PGA₂ analogues (**4a**-**4c**) indicated that the potency of isopeptidase inhibition for the series is I \gg Br \geq Cl.⁴ Additionally, iodo-PGA₂ was a more potent inhibitor of ubiquitin-specific isopeptidase activity compared to Δ^{12} -PGJ₂, but less potent than the PNGs. Thus, it was recognized that halogenation and electrophilicity do play a role in the ability of prosta-

^{(1) (}a) Mullally, J. E.; Moos, P. J.; Edes, K.; Fitzpatrick, F. A. J. Biol. Chem. **2001**, 276, 30366. (b) Mullally, J. E.; Fitzpatrick, F. A. Mol. Pharmacol. **2002**, 62, 351.

⁽²⁾ Masaaki, S.; Mori, M.; Niwa, T.; Hirata, R.; Furuta, K.; Ishikawa, T.; Noyori, R. J. Am. Chem. Soc. **1997**, 119, 2376.

⁽³⁾ Verbitski, S. M.; Mullally, J. E.; Fitzpatrick, F. A.; Ireland, C. M. J. *Med. Chem.* **2004**, *47*, 2062.

⁽⁴⁾ A paper thoroughly describing the pharmacology (outcome of the SAR studies and details of the biological assays data) will be published elsewhere.



glandins to inhibit ubiquitin isopeptidase activity but are not the only factors. These results suggested C-12 hydroxylation similar to PNGs may be necessary to achieve optimum activity. To test this hypothesis, a series of C-10 halogenated and C-12 oxygenated PGA₂ derivatives $(6a-6c)^4$ have been synthesized (Scheme 3).



The starting material for the synthesis, (15S)-prostaglandin-A₂ (PGA₂), was isolated in abundance $(5.7\% \text{ recovery})^5$ from the gorgonian *Plexaura homomalla*.⁶ The side-chain functional groups of PGA₂ (the terminal carboxylic acid at C-1 and the alcohol at C-15) were blocked as their methyl ester and acetate, respectively, using standard derivatization conditions (Scheme 1).⁷



⁽⁶⁾ Schneider, W. P.; Hamilton, R. D.; Rhuland, L. E. J. Am. Chem. Soc. 1972, 94, 2122.



(6d) X=H

The 15-*O*-acetyl-PGA₂ methyl ester (**2**) was treated with alkaline hydrogen peroxide at -15 °C to generate a mixture of C(10,11)- α , β -epoxides (**3**).⁸ The subsequent C-10 halide substitutions⁹ were achieved by regioselective ring opening of the C(10,11)-epoxide (**3**) with halide (Cl, Br, I) salts under mildly acidic conditions (Amberlyst 15¹⁰ or silica gel support¹¹). Similar transformations of epoxy cyclopentenones into α -haloenones have been shown to occur via the halohydrin intermediates, which are spontaneously dehydrated under acidic conditions to afford the vinylhalide products.¹⁰

Accordingly, the method of choice for the formation of vinyl chloride (**4a**) was LiCl/Amberlyst/CH₃CN system, where as the vinylbromide (**4b**) was formed by NaBr under otherwise identical conditions.¹² However, attempts to form the vinyl iodide (**4c**) under similar conditions (LiI or NaI/Amberlyst/CH₃CN) were unsatisfactory and resulted in the re-formation of the alkene (**2**). A subsequent literature survey of alternative methods of vinyliodide formation¹³ from epoxides revealed that silica gel could be employed as an efficient acid catalyst in the nucleophilic ring opening of epoxides under solvent-free conditions.¹¹ Accordingly, the solvent-free iodination of epoxide (**3**) was carried out using LiI supported on silica gel. The reaction proceeded smoothly

⁽⁷⁾ Schneider, W. P.; Bundy, G. L.; Lincoln, F. H.; Daniels, E. G.; Pike, J. E. J. Am. Chem. Soc. **1977**, 99, 1222.

⁽⁸⁾ $^1\!H$ NMR and HPLC analysis of crude mixture (3) showed no evidence of epimerization at C-8 during the epoxidation reaction.

 ^{(9) (}a) Dominguez, J. N.; Taddei, Â.; Cordero, M.; Blanca, I. J. J. Pharm. Sci. 1994, 83, 472. (b) Iguchi, K.; Kaneta, S.; Tsune, C.; Yamada, Y. Chem. Pharm. Bull. 1989, 37, 1173.

⁽¹⁰⁾ Righi, G.; Bovicelli, P.; Sperandio, A. Tetrahedron Lett. 1999, 40, 5889.

⁽¹¹⁾ Kotsuki, H.; Shimanouchi, T.; Ohshima, R.; Fujieara, S. *Tetrahedron* **1998**, *54*, 2709.

⁽¹²⁾ In a few instances during scale-up, the intermediate chloro- and bromohydrins were isolated. These halohydrins can be readily dehydrated to form the vinyhalides, by heating with glacial acetic acid on a steam bath for 2-5 h.

^{(13) (}a) Johnson, C. R.; Adams, J. P.; Braun, M. P.; Senanayake, C. B. W. *Tetrahedron Lett.* **1992**, *33*, 917. (b) Konaklieva, M. I.; Dahl, M. L.; Turos, E. *Tetrahedron Lett.* **1992**, *33*, 7093. (c) Palumbo, G.; Ferreri, C.; Caputo, R. *Tetrahedron Lett.* **1983**, *24*, 1307.

to give 54% of the vinyliodide (4c) along with 13% of the alkene byproduct (2).

The next task of the synthesis was the conversion of the C-10 halogenated PGA₂ series (**4a**-**4c**) to the corresponding C-12 hydroxylated derivatives (**6a**-**6c**). This novel series of C-12 hydroxylated derivatives was believed to be accessible via allylic oxidation mediated by selenium dioxide¹⁴ because the C-12 position of PGA₂ is bisallylic to C-10 and C-13 double bonds (Scheme 2). However, the likely formation of byproducts due to multiple potential oxidation sites was foreseen as a possible drawback of this method.

To test the feasibility of the allylic oxidation reaction, PGA_2 -acetate methyl ester (2) was employed. As speculated, compound 2 with SeO₂ gave several oxidation byproducts. One major byproduct was identified as the conjugated diene (2a) possibly arising from allylic hydroxylation at C-7 followed by dehydration (Scheme 2). Shorter reaction times, lower reflux temperatures (95% EtOH), or reduced amounts of the oxidant did not facilitate formation of the desired tertiary alcohol product (6).

It was now evident that in order to selectively hydroxylate the bisallylic position (C-12) of PGA₂ and suppress byproduct formation during SeO₂ oxidation, the other allylic positions (C-4 and C-7) needed to be removed or masked in some manner. This was achieved by selective epoxidation of the C(5,6) double bond in **4** using *m*-CPBA (Scheme 3). The resulting C(5,6)- α , β -epoxide mixture (**5**) was treated as an intermediate and used without further purification for the SeO₂ oxidation. As anticipated, allylic hydroxylation at the C-12 position proceeded smoothly and was accompanied by C(5,6)-epoxide ring opening to yield the novel series of prostaglandin-A₂ analogues (**6a**-**6c**)¹⁵ in moderate yields (Scheme 3).

The relative and absolute configurations of the five stereocenters (C-5/6/8/12/15) in (**6a**-**6d**) were deduced as follows. The absolute stereochemistry of C-8 was assumed to be (*R*), based on literature precedent for coral-derived PGA₂.¹⁶ The assignment of the 15(*S*) configuration of **6** was based on the application of Mosher's method¹⁷ to (*S*)- and (*R*)-MPA derivatives of PGA₂-methyl ester.¹⁸ The *trans* (*R*,*R*) relationship between the two side chains (C-8/C-12) was established based on a strong NOESY cross-peak observed between H-8 and H-13. Subsequent molecular modeling studies with energy-minimized conformations for each of the C-12 epimers supported this observation. The *trans* (*8R*,12*R*) relationship was further corroborated by heteronuclear coupling constant analysis (²J_{H8/C12} = small)¹⁹ and

dihedral angle measurements (-135.7°) for the (8R, 12R) diastereomer of **6**.²⁰

The *cis* orientation of the C(5,6) epoxide mixture [(5S,6R)and (5R, 6S) diastereomers] in 5 was supported by the homonuclear coupling constant value of ${}^{3}J_{\text{H5/H6}} = 4.3 \text{ Hz.}^{21}$ It was presumed that the C(5,6) diol in 6, generated during the allylic oxidation (SeO₂/dioxane/H₂O/reflux) of 5, occurs via nucleophilic opening of the C(5,6) epoxide by water. Accordingly, nonregioselective attack of water on the (5S, 6R)and (5R,6S) cis-epoxides in 5 would generate two secondary alcohol diastereomers, (5R, 6R) and (5S, 6S). This was supported by the ¹H NMR spectra of (6a-6d), which showed doubling of resonances due to the formation of two diastereomers in a \sim 1:1 ratio. Accordingly, HPLC analysis of **6d** showed two peaks eluting at 11.5 (7) and 13 min (8). The two peaks (7 and 8) were separated²² and were employed for absolute stereochemical assignments of the C(5,6) centers by J-based analysis,¹⁹ and Mosher's method¹⁷ in conjunction with molecular modeling studies (Figure 2).



Figure 2. C(5,6)-Diol diastereomers of 6d.

Initial attempts focused on utilizing *J*-based analysis¹⁹ to relate the configuration of the vicinal diol segment C(5,6) to that of the known absolute configuration at C-8. However, diastereomer **7** proved undesirable for *J* analysis due to overlap of resonances for H-5 and H-6 (δ 3.51–3.57, m, 2H) as well as for H-7_h and H-7₁ (δ 1.98, m, 2H). Therefore, the viable alternative was to attempt the *J*-based analysis on the diastereomer **8**.

For diastereomer **8**, sufficient chemical shift dispersion of all proton resonances was observed in CDCl₃ at 0 °C. While all of the couplings required for *J* analysis were determined successfully (Table 1), the information failed to fit any of the typical rotamers with certainty.¹⁹

The reason for the occurrence of atypical coupling values in **8** could be explained based on a molecular modeling study, which revealed that the cyclopentenone carbonyl (C-9) is involved in an intramolecular H-bond with C-6(OH) adopting a seven-membered cyclic conformation (Figure 3). The measured J values and NOESY data were in accordance with such a seven-membered conformation.

^{(14) (}a) Sharpless, K. B.; Lauer, R. F. J. Am. Chem. Soc. 1972, 94, 7154.
(b) Groszek, G.; Tyrlik-K, A.; Wicha, J. Tetrahedron 1989, 45, 2223.

⁽¹⁵⁾ NMR data were recorded at 0 °C. Compounds stored at rt tend to decompose slowly over 3-5 days.
(16) (a) Baker, B. J.; Okuda, R. Y.; Yu, P. T. K.; Scheuer, P. J. J. Am.

^{(10) (}a) Baket, B. J.; Okuda, R. 1.; 10, P. 1. K.; Scheder, P. J. J. Am. Chem. Soc. **1985**, 107, 2976. (b) Weinheimer, A. J.; Spraggins, R. L. Tetrahedron Lett. **1969**, 10, 5185.

^{(17) (}a) Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. J. Am. Chem. Soc. **1991**, 113, 4092. (b) Seco, J. M.; Quiñoà, E.; Riguera, R. Tetrahedron: Asymmetry **2001**, 12, 2915.

⁽¹⁸⁾ An attempt to determine the absolute stereochemistry by a variable temperature ¹H NMR experiment with a single MPA derivative, (15*R*)-MPA, in CDCl₃ at -1 and 25 °C was not successful: Latypov, S. K.; Seco, J. M.; Quiñoà, E.; Riguera, R. *J. Am. Chem. Soc.* **1998**, *120*, 877.

⁽¹⁹⁾ Matsumori, N.; Kaneno, D.; Murata, M.; Nakamura, H.; Tachibaba, K. J. Org. Chem. **1999**, 64, 866.

⁽²⁰⁾ Contreras, R. H.; Peralta, J. E. Prog. NMR Spectrosc. 2000, 37, 321.

⁽²¹⁾ Due to the overlap of H-5 and H-6, a combination of ${}^{1}\text{H}$ NMR, HOMODEC, and 1D TOCSY experiments was employed to extract the coupling constants with certainty.

⁽²²⁾ See experimental section for details of the separation procedure.

Table 1. ^{2,3}*J* Values of **8** for the C-6/C-7 Segment Measured in $CDCl_3$

$^{2,3}\!J\mathrm{values}^a$	$J\left(\mathrm{Hz}\right)$	${\it classification}^b$
${}^{3}J$ (H-6, H-7 _h) ${}^{3}J$ (H-6, H-7 _l) ${}^{3}J$ (H-6, C-8) ${}^{3}J$ (C-5, H-7 _h) ${}^{3}J$ (C-5, H-7 _l) ${}^{2}J$ (C-6, H-7 _h)	+6.3 +6.3 +2.2 +4.4 +2.2 -4.3	medium medium small medium small medium
${}^{2}J$ (C-6, H-7 ₁)	-1.1	small

^{*a*} H-7_h and H-7_l represent the high- and low-field H-7 protons, respectively. ^{*b*} Classification for magnitude of the coupling constants; values between large and small are regarded as medium.¹⁹

Having failed to unequivocally determine the C(5,6) stereochemistry by *J*-based analysis, Mosher's method was attempted on **8**. Accordingly, reaction of **8** with (*R*)- and (*S*)-MPA acids²³ was expected to yield the corresponding C(5,6)-bis-MPA esters. However, ¹H NMR data confirmed that the C-5 MPA derivative was the sole product. Comparison of the chemical shift differences ($\Delta \delta = \delta_R - \delta_S$) in the ¹H NMR spectra of the C-5 (*R*)- and (*S*)-MPA derivatives of **8** at 0 °C indicated the absolute configuration was (*R*) (Figure 4).

Thus, the absolute configuration of all five stereocenters (C-5/6/8/12/15) in **8** was established as (5R,6R,8R,12R) and



Figure 3. The MM2/AM1-optimized model for 8.



Figure 4. The $\Delta \delta$ values for **8** ($\Delta \delta = \delta_R - \delta_S$, 500 MHz, CDCl₃, 0 °C).

(15*S*). Consequently, the absolute configuration of the diol diastereomer **7** was assigned as (5S,6S,8R,12R) and (15S). The absolute configuration assignments of **7** and **8** were in good agreement with their ¹H NMR data, coupling constant analysis, and molecular modeling studies employing MM2 and AM1.²⁴

Acknowledgment. The authors wish to acknowledge Dr. Joseph R. Pawlik (University of North Carolina at Wilmington) for collection of the gorgonian. Dr. P. Krishna at the University of Utah Mass Spectrometry and Proteomics Core Facility is acknowledged for performing the LRESIMS experiments. The HRESIMS data were acquired at the University of Iowa High Resolution Mass Spectrometry Facility. This project was funded by NIH Grant CA 36622.

Note Added after ASAP Publication. There were two errors in Table 1 in the version published ASAP April 15, 2006; the corrected version was published April 21, 2006.

Supporting Information Available: Experimental details and spectroscopic data for synthetic compounds 2-8. This material is available free of charge via the Internet at http://pubs.acs.org.

OL0606583

^{(23) 2} equiv of MPA per OH was employed. Reaction was carried out at room temperature for \sim 16 h, until complete conversion to the product, as indicated by TLC and NMR.

⁽²⁴⁾ CS Chem3D Pro; ChembridgeSoft Corporation: Cambridge, MA.